# EFFECTS OF STARVATION AND ALCOHOL CONSUMPTION ON SOME METABOLIC ASPECTS IN MALE ALBINO RATS .

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#### Abstract

This study was conducted to examine the effects of starvation in presence or absence of alcohol consumption on some metabolic pathways in rats. The results indicated that starvation decreased blood glucose, total proteins and albumin concentrations, whereas insulin and glucagon increased. On the other hand, serum enzymes including aspartate aminotransferase (AST), alanine aminotransferase (ALT) and lactate dehydrogenase (LDH) activities were increased in response to complete starvation, but alkaline phosphatase (ALP) and gamma-glultamyltransferase (GGT) activities decreased, however, acid phosphatase was approximately unaltered. Ethanol was found to decrease serum (ALT) and (LDH) activities, while serum (GGT) was increased.

Hepatic glycogen was found to decrease in starved and alcoholic fasted rats. Aminotransferases of the liver were decreased in response to starvation. Liver succinic dehydrogenase activity increased at 1 and 3 days of fasting, but decreased insignificantly at the 5 th day of starvation.

Ethanol induced an increase in hepatic ALT activity at the 5 <u>th</u> day of starvation, but it decreased glycogen content, AST and succinic dehydrogenase activities in fasted rat liver. The present findings support the suggestion that starvation disturbs most of the mabolic pathways while alcohol intake of starved animals exacerbates such a disturbance.

#### INTRODUCTION

It was reported that starvation causes many alterations in animal physiology and biochemistry, including the body weight (Chandrasekharan, 1969 and Inoue *et al.*, 1993), blood glucose level (Brooke, 1981), hepatic and muscle glycogen (Cleader and Robert, 1992), plasma glucagon like immunoreactivity (Takahashi *et al.*, 1992), serum and liver lipids and proteins (Mendez and Menchu, 1966 and Sassoon *et al.*, 1966), hepatic glucose- 6- phosphate dehydrogenase (McDonald and Johnson, 1965), hepatic ALT, liver AST (Stielau *et al.*, 1965) and arterial alkaline phosphatase activities (Zemplenyi, 1962).

These changes may also be affected in case of ethanol intake after starvation (Forsander *et al.*, 1965). This paper investigates the influence of starvation on different metabolic parameters in rat liver and serum with special reference to ethanol interaction.

#### MATERIAL AND METHODS

## Animals:

Male albino rats (*Rattus norvegicus*), purchased from the Egyptian organization for Biological and vaccine production, A.R.E., weighing about  $100 \pm 20g$ , were used as experimental animals throughout the present work. Animals were housed in groups in plastic cages.

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#### Experimental Design:

## Animals were segregated into 3 groups as follows:

1- The first group received balanced diet and tap water ad-libitum and served as control.

2- The animals of the second group were starved and received only tap water ad-libitum.

3- Animals of the third group were starved and received 20% of ethanol in tap water *ad-libitum*.

Animals of different groups were sacrified after 1, 3 and 5 days of starvation.

#### **Collection of Serum Samples:**

Blood samples were received in conical glass centrifuge tubes and were put in refrigerator till clotting. Sera were then separated using cooling centrifuge at 1000 g for 10 min. and kept at-20°C till analyses.

## Preparation of Tissue Homogenates:

After decapitation, animals were dissected and livers were immediately excised, blotted with filter paper and frozen in a deep freezer at-20°C till further experimental procedures. Accurately weighted hepatic portions were homogenized in an ice bath using glass homogenizer to give a final dilution of 10% tissue homogenate which is further diluted in order to be convenient for determinations of different parameters.

#### **Biochemical Analysis:**

Serum total protein and albumin contents were determined using an automatic multiparameter apparatus, ASTRA<sup>8</sup> synchron AS clinical system.

Blood glucose concentration was determined colourimetrically using dimension <sup>®</sup> ES machine, The Liver Institute, Menoufia University, Egypt. Serum glucagon concentration was carried out by the method of Harris *et al.* (1979) using radioimmunoassay kits of Biomedicals Inc. Company, USA. Insulin was determined in serum according to the radioimmunoassay method of Wilson and Miles, (1977), using kits of SORIN Biomedica Company, Italy.

Serum aspartate aminotransferase (AST) [Ec. 2.6.1.1], alanine aminotransferase (ALT) [Ec. 2.6.1.2], alkaline phosphatase (ALP) [EC. 3.1.3.1], acid phosphatase [Ec. 3.1.3.2] and gamma-glutamyltransferase (GGT) [Ec. 2.3.2.2] activities were determined using an automatic multiparameter apparatus, ASTRA<sup>8</sup> synchron AS clinical system. Lactate dehydrogenase (LDH) [Ec. 1.1.1.2] activity in serum was determined using kits of human gesllschaft flr biochemica Ind diagnostica mbh. Hepatic glycogen contents were quantifid colourimetrically by the method of Seifter *et al.*, (1950). Hepatic AST and ALT activities were measured by the method of Reitman and Frankel (1957), using commercial kits of Al-Gomhoria Co., Egypt Hepatic succinic dehydrogenase activity was measured colourimetrically using the method of Kun and Abood, (1949).

#### **Statistical Analysis:**

Data are presented as means  $\pm$  standared deviation (S.D.) in Tables. Student's t-test was used to evaluate statistical significances of results according to Hine and Wetherill, (1975).

## RESULTS

## Blood parameters

## Total Proteins:

As represented in (Table. 1), it can be noticed that serup total protein content decreased after 1, 3 and 5 days of starvation by 7.31, 8.68 and 10.97%, respectivily, while starvation with ethanol intake resulted in increased these depression magnitudes (Table. 2)

#### Albumin:

Albumin decreased significantly in starved rats (Table. 1). Similar results were observed with ethanol intoxication (Table. 2).

#### Glucose:

It was found that serum glucose level decreased highly significantly by the ratios: 52.46, 52.67 and 55.14 at 1,3 and 5 days of starvation (Table. 1), similar results were observed after alcohol abuse of starved rats (Table.2).

#### Glucagon:0

This parameter showed highly significant increases in serum after starvation, which were amounted to 35.55, 22.42 and 96.43% of their respective control values at 1,3 and 5 days, respectively (Table. 1)

On the other hand, ethanol intake causes blood glucagon level to increase highly significantly by 27.92, 77.83 and 197.79% after 1,3 and 5 days of treatment, respectively (Table. 2).

## Insulin:

Radioimmunoassay analysis of insulin has resulted in a significant increase in serum insulin content of starved rats at 3 rd day of experiment, and a highly significant (P < 0.01) elevation of 101.11% at 5 th day of starvation (Table. 1).

Values of serum insulin in starved rats after ethanol intoxication (Table, 2) exhibited highly significant increases amounted to 56.66 and 53.30% at 1 and 5 days of treatment, respectively.

#### Aminotransferases:

Serum ALT and AST activities were noticed to increase at different periods of starvation, but insignificant depression was observed in serum AST activity at 1 st day of treatment (Table 3).

On the other hand, these enzyme activities were noticed to increase at different periods of starvation of ethanolic rats however, a highly significant decrease in hepatic ALT activity was observed at 5 days of treatment (Table. 4).

#### Alkaline phosphatase:

This enzyme activity was decreased highly significantly with magnitudes of 31.30 and 41.29% at 3 and 5 days of starvation (Table. 3), while it diminshed at 1,3 and 5 day of ethanol intake in completely starved rats by the ratios: 44.33, 46.52 and 48.94%, respectively (Table. 4).

#### Acid phosphatase:

As shown in table (3), the activity of acid phosphatase was decreased at 1 and 3 days of starvation while it increased at the 5 th day of fasting (Table. 3). On the other hand this parameter decreased at the 1 st day of starvation of ethanolic rats, but increased at 3 and 5 days of treatment (Table. 4).

## Gamma glutamyl transferase:

The changes in serum GGT activities following starvation were presented in table (3). It was found that GGT activities were decreased highly significantly at different periods of starvation.

Starvation of ethanol-treated rats showed highly significant decreases in the activity of this enzyme at 1 and 3 days of treatment by the values 43.55 and 38.99%, respectively (Table. 4).

## Lactate dehydrogenase:

This enzyme showed highly significant (P < 0.01) elevations amounted to 70.92 and 43.59% at 1 and 3 days of starvation (Table. 3), while it increased only at 1 st day of ethanol intake of starved rats, then it decreased insignificantly at 3 and 5 days of treatment.

## Liver parameters:

## Glycogen:

Hepatic glycogen content were found to be greatly decreased under the influence of starvation or fasting with alcohol abuse (Tables 5 and 6).

#### Transaminases:

Aminotransferase activities in liver of starved rats were decreased highly significantly after all periods of the experiment (Table. 5).

Similarly, hepatic aminotransferases decreased in response to starvation and alcohol drinking at different periods of treatment but with an exception at day 5 of treatment where liver ALT activity was found to increase insignificantly (Table. 6).

#### Succinic dehydrogenase:

Starvation induced to increases in the activity of this enzyme at 1 and 3 days of fasting while it caused hepatic succinic dehydrogenase activity to decrease insignificantly at the 5  $\underline{\text{th}}$  day of fasting (Table 5).

Hepatic succinic dehydrogenase activity in starved rats was found to decrease highly significantly by the magnitudes: 42.72, 47.39 and 44.06% at 1,3 and 5 days of ethanol treatment, respectively (Teble. 6).

#### DISCUSSION

The results showed that blood glucose and hepatic glycogen contents were decreased after starvation with or without alcohol abuse. These results are in agreement with those reported by Morata *et al.* (1982), who referred to the decrease in gluconeogenesis process and to the enhancement of glycogenolysis in the liver because of the utilization of liver glycogen as a source of blood glucose during the initial period.

The depression in blood glucose during starvation is associated with an increase in serum insulin and glucagon content that was in accordance with the opinion that fasting facilitates glucose utilization (Unger *et al.*, 1963 and Tsutomu *et al.*, 1992). Pico *et al.* (1991) reported that starvation increases red cells amino acids uptake; this may partially interpreter the decrease in serum total prateins. Our results for ethanol intoxicated starved rats are in agreement with those reported by Baraona and Lieber, (1982) and Brassinne, (1979).

Ethanol was reported to increase serum ALT activity (Piedras *et al.*, 1987 and Shalan(M), 1996); where alcohol may exert its effect through alterations it induces on synthesis in the endoplasmic reticulum, intracellular translocation or possibility of solubilization at the site of plasma membrane, hence increasing the level of serum enzymes especially membrane-bound enzymes such as GGT, ALP and cytosolic enzymes such as ADH, LDH and aminotransferase enzymes (ALT and AST).

Forsander *et al.* (1965) reported that ethanol treatment of starved rats decreased the pyruvate concentration which can be referenced for the decrease in ALT and LDH activities at 5 days of ethanolic starved rats. Moderate acohol consumption resulted in small but significant elevation in GGT and urate while AST, folate and fibrinogen showed no change (Lieber, 1984 and Pikaar *et al.*, 1987).

Shalan(A), (1995) showed that following long-term alcohol intake, striking increase of serum ALP activity was observed in male albino rats fed laboratory balanced diet. However Goz *et al.* (1983) and Nishimura and Teschke (1982) demonestrated that ethanol didn't affect ALP activity. The increased serum activity of GGT at day 5 of ethanol intoxication in starved rats may result from the induction of this enzyme in the liver and/ or due to damage of hepatocytes and other organ tissues known to be rich in this enzyme such as renal tissues (Jacquemin *et al.*, 1990).

On the other hand, it was recorded that hepatic alkaline phosphatase and aminotransferase (ALT and AST) activities decreased insignificantly in response to chronic alcohol intake (Teschke *et al.*, 1983; Yamada *et al.*, 1985 and Adel-Raheem *et al.*, 1990).

The fact that present findings showed that starvation causes an increase in hepatic succinic dehydrogenase activity at 1 and 3 days, may contribute to increased rate of citric acid cycle and glycogenolytic process (Bayomy and Taie 1992). However, hepatic succinic dehydrogenase was reported to decrease in response to chronic alcohol intoxication (Shalan(M), 1996).

In conclusion the present data indicates that alcohol eggravates metabalic disturbance induced in starved rats as indicated by the chosen parameters measured in both serum and liver homogenate.

#### RESFERENCES

- Abdel-Raheem, K.; El-Mosallamy, N.; Bayomy, M.F.F., Mohamed, A. and El-Elaimy, I. (1990). Metabolic responses to alcohol intoxication. II. Amino acid and protein metabolism. Proc. Zool. Soc., 18:253-268.
- Baraona, E. and Lieber, C.S. (1982). Effect of ethanol on serum proteins. Ann. Rev. Med., 33: 281-292.
- Bayomy, M.F.F. and Taie, H.T. (1992). Effects of fasting, refeeding and restricted feeding on the young rabbit. II. Influence on fuel metabolism. Egypt. J. Rabbit Sci., 2:49-60.

- Brassinne, A. (1979). Effect of ethanol on plasma protein shedding in the human stomach. Digest. Dis. Sci., 24:44-47.
- Brooke, J.D. (1981). Lowered blood glucose with human fasting, exercise and carbohydrate diet. Nutr. Rep. Int., 24:1279-1286.
- Chandrasekharan, N. (1969). Effect of starvation and subsequent refeeding on body and liver weights. Nutr. Dieta., 11:53-59.
- Cleader, P.C. and Robert, G. (1992). Heterogeneity of glycogen synthesis upon refeeding following starvation. Int. J. Biochem., 24:71-78.
- Forsander, O.A.; Raiha, N., Salaspuro, M. and Maenpaa, P. (1965). Influence of ethanol on the liver metabolism of fed and starved rats. Biochem. J., 94:259-265.
- Goz, B.; Stawe, A.C. and Townsend, A.J. (1983). Effect of ethanol on alkaline phosphatase activity in Hela cells. Alcohol. Clin. Exp. Res., 7:176-179.
- Harris, V.; Maclian, A. and Vali, M. (1979). Radioimmunoassay of glucagon. In: Methods of Hormone Radioimmunoassay. [Eds. A., Jaffe and C. Behrman], P. 643, Academic press, New York.
- Hine, J. and Wetherill, G.B. (1975). A programmed text in statistics. Book 3. The t-text X<sup>2</sup> Goodness of fit. Chapman and Hill, London.
- Inoue, E. A.; Yoshifum, A. S.; John, P.G. and Phyllis, J.S. (1993). Effect of glutamine- supplemented total parenteral nutrition on recovery of the small intestine after starvation atrophy. J. Parenter. Enteral. Nutr., 17:165-170.
- Jacquemin, E.; Bulle, F.; Bernaudin, J.F.; Wellman, M.; Hugan, R.N.; Guellaren, G. and Hadchouel, M. (1990). Pattern of expression of γ-glutamyltranspeptidase in rat liver and kidney during development. Study by immunochemistry and in situ hyperdization. J. Pediat. Gastroentrol. Nut., 11:89-95.
- Kun, E. and Abood, L.G. (1949). Colourimetric estimation of succinic dehydrogenase by triphenyltetrazolium chloride. Science, 109:144-146.
- Lieber, C.S. (1984). Alcohol and the liver, 1984 update. Hepatology, 4:1243-1260.
- McDonald, B.E. and Johnson, B.C. (1965). Metabolic response to realimentation following chronic starvation in the adult male rat. J. Nutrit., 87:161-167.
- Mendez, J. and Menchu, M.T. (1966). Effect of dietary protein level prior to acute starvation on serum proteins in the rat. J. Nutrit., 88:365-369.
- Morata, P.; Vargas, A.M.; Sanchez- Medina, F.; Garcia, M.; Cardenete, G. and Zamora, S. (1982). Evolution of gluconeogenic enzyme activities during starvation in liver and kidney of the rainbow trout (*Salmo gairdner*). Comp. Biochem., Physiol. B. Comp. Biochem., 71:65-70.
- Nishimura, M. and Teschke, R. (1982). Effect of chronic alcohol consumption on the activities of liver plasma enzymes: gamma- glutamyltransferase, alkaline phosphatase and 5'-nucleotidase. Biochem. Pharmacol., 31:377-381.
- Pico, C.; Pona, A. and Palou, A. (1991). Start term starvation-induced changes in the kinetic parameters of rat red cell L-alanine and glycine uptake. Biochem. Int., 25:1095-1103.
- Piedras, J.; Iagulirre, R.; Lisker, M.; Frias, J.; Karpovitch, X. L. and Cordova, M.S. (1987). Hepatic iron stores and serum ferritin level in alcoholic and nonalcoholic liver disease. Lab. Res. Invest. Clin. (Mex.)., 39:343-347.

- Pikaar, N.A.; Wedel, M.; Vander Beek, E.J.; Wim Van Dokkun, Kempen, H.M.; Kluft, C.; Ockhuizen, T. and Hermus, R.J.J. (1987). Effects of moderate alcohol consumption on platelet aggregation, fibrinolysis, and blood lipids. Metabolism, 36:538-543.
- Reitman, S. and Frankel, S. (1957). A colourimetric method for determination of serum glutamic- oxaloacetic and glutamic- pyruvic transaminase. Am. J. Clin. Path., 28:56-63.
- Sassoon, H.F.; Jonson, B.C. and Maser, K. (1966). Dietary carbohydrates and possible pre-lesion biochemical changes in the aortas of adult male rats. J. Nutrit., 90:275-283.
- Seifter, S.; Daytan, S.; Novic, B. and Muntwylers, E. (1950). The estimation of glycogen with the anthrone reagent. Arch. Biochem., 25:191-200.
- Shalan, A.G. (1995). Physiological and biochemical studies of effects of ethyl alcohol on liver and testes of the growing rat. M.Sc. Thesis, Fac. Sci., Menoufia Univ., Egypt.
- Shalan, M.G. (1996). Biochemical studies of effects of alcohol consumption on fat and carbohydrate metabolism in rats fed different levels of proteins. M.Sc. Thesis, Fac. Sci., Menoufia Univ., Egypt.
- Stielau, W.J.; Freedland, R.A. and Meyer, J.H. (1965). Effects of B-vitamin deficiencies and of starvation on liver enzyme activities of growing rats. J. Nutrit., 87: 109-116.
- Takahashi, J.I.; Noma, Y.; Yoshimoto, S.; Fujita, C. and Shima, K. (1992). Decrease in plasma GLP-1 immunoreactivity in starved rats. Diabetes Res. Clin. Pract., 15:205-212.
- Teschke, R.; Neuefeind, M.; Nishimura, M. and Strohmeyer, G. (1983). Hepatic gamma-glutamyltransferase activity in alcoholic fatty liver: Comparison with other liver enzymes in man and rats. Gut, 24:625-630.
- Tsutomu, O.; Manalu, N.; Motoyoshi, K.; Toshitaka, K. and Yoshio, I. (1992). Starvation induced change of glucose utilization in the rat skeletal muscle. Jikeikai. Med. J., 39:269-274.
- Unger, R.H.; Eisentraut, A.M. and Madison, L.L. (1963). The effects of total starvation upon the levels of circulating glucagon and insulin in man. J. Clin. Invest., 42:1031-1037.
- Wilson, M.A. and Miles, L.S.M. (1977). Radioimmunoassay of insulin. In: Handbook of radioimmunoassay. [Ed. G.E. Abraham], P. 275. M. Dekker Inc. Press, New York.
- Yamada, S.; Wilson, J.S. and Lieber, C.S. (1985). Effects of alcohol and diet on hepatic and serum gamma-glutamyltranspeptidase activities. J. Nutrit., 115:1285-1290.
- Zemplenyi, T. (1962). Enzymes of the arterial wall. J. Atheroscler. Res., 2:2-7.

			٦	Duration of Starvation							
		Control		l day		3 days		5 days			
		Mean ± S.D	).	Mean ± S.D.	% diff.	Mean ± S.D.	% diff.	Mean ± S.D.	% dif:(.		
Glucose	а	97.2	1	46.2	**	46	**	43.6	**		
•	_	± 6.05		± 7.69	-52.46	±1.58	-52.67	± 2.70	-55.14		
Glucagon	a	188.62	2	255.68	**	230.92	**	370.52	**		
	-	± 13.4		± 20.28	35.55	± 16.25	22.42	± 16.48	<b>96.4</b> 3		
Insulin	a	10.73	3	13.25		13.27	*	21.58	**		
	- 1	± 0.55		± 2.48	23.48	± 1.90	23.6	± 2.67	101.11		
Total Protein	а	6.56	4	6.08	*	5,99	**	5.84	**		
	<u> </u>	± 0.24		± 0.015	-7.31	± 0.03	-8.68	± 0.21	-10.97		
Albumin	a	3.46	5	1.87	**	1.17	**	1.13	**		
	~	± 0.23		± 0.02	-45.95	± 0.015	-66.18	± 0.011	-67.34		

Table (1): Effect of starvation on some blood metabolites of rats.

a: n = 5.ı : mg/dl. 2 : Pg/ml.

Highly significant (P < 0.01). 4: gm/dl.

s : gm/dl.

\* = Significant ( P < 0.05).

Table (2): Effect of ethanol intake on some blood metabolites in starved rats.

			Duration of Starvation							
		Control	1 ds	ıy	3 d	lays	5 days			
		Mean ± S.D.	Mean ± S.D.	% diff.	Mean ± S.D.	% diff.	Mean ± S.D.	% diff.		
Glucose	a	97.2 ± 6.05	54.4 ± 3.36	**	52.2 ± 2.86	** -46.29	42 ± 4.63	-56.79		
Glucagon	a	188.62 <sup>2</sup> ±13.4	241.30 ±29.23	** 27.92	335.43 ± 29.24	** 77.83	561.70 ± 30.06	** 197.79		
Insulin	a	$10.73  3 \\ \pm 0.55$	16.81 ± 2.42	** 56.66	13.05 ± 2.17	* 21.62	16.45 ± 3.07	** 53.30		
Total Protein	a	6.56 4 ± 0.24	5.42 ± 0.31	** -17.37	4.90 ± 0.16	** -25.3	4.70 ± 0.16	** -28.35		
Albumin	a	3.46 5 ± 0.23	1.16 ± 0.03	** -66.47	1.04 ± 0.02	** -69.94	0.974 ± 0.011	** -71.96		

3 : μu/ml. a:n=5.4 : gm/dl. ı:mg/dl.

2 : Pg/ml. s:gm/dl. \*\* = Highly significant (P < 0.01).</li>
\* = Significant (P < 0.05).</li>

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# Effects of starvation and alcohol consumption.....

		Duration of Starvation							
	Control	1 day		3 days		5 days			
	Mean ± S.D.	Mean ± S.D.	% diff.	Mean ± S.D.	% diff.	Mean ± S.D.	% diff.		
AST	183.6 ± 15.94	171.6 ± 2.41	- 6.99	186 ± 3.67	1.307	194.4 ± 2.70	5.88		
ALT a	39.8 <sup>2</sup> ± 1.92	58.80 ± 1.92	* 47.73	47.4 ± 3.64	* 19.09	43 ± 2.73	8.04		
ALP a	$164.2 \xrightarrow{3}{\pm} 12.67$	146.4 ± 12.62	-10.84	112.8 ± 3.11	* -31.30	96.4 ± 2.4	* -41.29		
Acid phosphatase a	1.17 4 ± 0.13	0.734 ± 0.09	-37.31	1.14 ± 0.08	-2.81	1.27 <u>+</u> 0.11	8.45		
GGT a	27.21 <sup>5</sup> ± 2.00	11.53 ± 0.04	* -57.66	13.72 ± 0.23	* -49.65_	20.48 ± 0.33	* -24,73		
Lactate a dehydrogenase	775.8 6 ± 31.32	1326 ± 91.14	* 70.92	1114 ± 85.32	* 43.59	879 ±158.11	13.03		

Table (3): Effect of starvation on some blood enzymes of rats

a: n = 5.3 : Iu/L. 4 : U/L. i : Iu/L.

6 : U/L.

2 : Iu/L . 5: U/L. \* = Highly significant (P < 0.01).

Table (4): Effect of ethanol intake on some blood enzymes of starved rats.

		Duration of Starvation						
	Control	1 day		3 days		5 days		
	Mean ±	Mean ±	% diff.	Mean ±	% diff.	Mean ±	% diff.	
	S.D.	S.D.		S.D.		S.D.		
AST	a 183.6 1	185.6		197.2		207.2	*	
•	± 15.94	± 2.19	1.08	± 2.16	7.40	± 5.93	12.85	
ALT	a 39.8 2	51.6	**	40.4	· · · · · · · · · · · · · · · · · · ·	33.4	**	
	± 1.92	± 1.81	29.64	± 1.14	1.50	± 1.14	-16.08	
ALP	164.2 3	91.40	**	87.8	**	83.83	**	
	± 12.67	± 3.50	-44.33	± 1.30	-46.52	± 2.22	-48.94	
Acid phosphatase	1.17 4	0.35	**	1.24		1.73	**	
•••	± 0.13	± 0.079	-70.11	± 0.07	5.89	± 0.08	48.42	
GGT	27,21 5	15.36	**	16.60	**	28.91		
	± 2.00	± 0.24	-43.55	± 0.20	-38.99	± 0.31	6.24	
Lactate	775.8	1242.2	**	687		706		
dehydrogenase	± 31.32	± 84.65	60.11	±114.01	-11.44	± 119.07	-8.99	

a: n = 5.3 : Iu/L.

1 : Iu/L. 4:U/L.

2 : Iu/L.

5: U/L.

6 : U/L. \*\* = Highly significant (P < 0.01). \* = Significant (P < 0.05).

Table (5): Effect of starvation on hepatic glycogen content, ALT, AST and succinic dehydrogenase activities of rats.

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			Duration of Starvation							
	L	Control Mean ± S.D.	Control 1 day		3 d	lays	5 days			
			Mean ± S.D.	% diff.	Mean ± S.D.	% diff.	Mean ± S.D.	% diff.		
Glycogen	а	35.31 I ± 5.54	22.92 ± 1.89	** -35.08	11.00 ± 1.54	** -63.18	21.12 ± 1.19	** -40.18		
ALT	a	11.47 <sup>2</sup> ± 0.49	10.16 ± 0.28	**	9.09 ± 0.33	** -20.74	$9.81 \pm 0.104$	**		
AST	a	21.42 3 + 2.66	6.86 ± 0.25	** -68.16	5.81 ± 0.17	-72.87	2.97 ± 0.125	** -76.79		
Succinic dehydrogenase	a	1.65 4 ± 0.035	1.829 ± 0.065	** 10.85	1.736 ± 0.065	* 5.21	1.619 ± 0.036	-1.87		

a : n ≈ 5.

1 : mg/gm wt. tissue. 2 : U/gm wt. tissue.

3 : U/gm wt. tissue .

4: ug of dye reduced/gm of tissue/ 10 min.
\*\* = Highly significant (P < 0.01).</li>
\* = Significant (P < 0.05).</li>

Table (6):	Effect of ethan	ol intake on hepatic glycogen content, ALT, AST and
	succinic dehydr	ogenase activities of starved rats.

	T		Duration of Starvation							
	L	Control	Control 1 da		y 3 day		50	days		
		Mean ± S.D.	Meau ± S.D.	% diff.	Mean ± S.D.	% diff.	Mean ± S.D.	% diff.		
Glycogen	a	35.31 1 ± 5.54	20.78 ± 0.69	** -4].14	22.25 ± 0.60	** - 36.98	27.14 ± 0.46	* - 23,25		
ALT	a	11.47 2 <u>+</u> 0.49	10.42 ± 0.21	** -9.15	11.2 ± 0.29	-2.35	11.50 ± 0.16	0.261		
AST	a	21.42 3 ± 2.66	6.38 ± 0.29	-70.21	5.24 ± 0.17	** -75.53	4.58 ± 0.16	** -78,86		
Succinic dehydrogenase	a	1.65 4 ± 0.035	0.945 ± 0.061	** -42.72	0.868 ± 0.054	** -47.39	0.923 ± 0.066	** -44.06		

a:n=5.

1 : mg/gm wt. tissue.

2 : U/gm wt. tissue.

3 : U/gm wt. tissue .

4 : ug of dye reduced/gm of tissue/ 10 min. \*\* = Highly significant (P < 0.01). \* = Significant (P < 0.05).

Effects of starvation and alcohol consumption.....

تأثير التجويع وإستهلاك الكحول على بعض المؤشرات الأيضيه في ذكور الجرذان البيضاء.

محمد فتحى بيومى، 
 محمد جابر محمد شعلان، 
 عادل جابر محمد شعلان
 كلية العلوم – جامعة المنوفية – مصر
 كلية التربية – جامعة فناة السويس – مصر

# الملخص العربي

تم فى هذا البحث دراسة تأثير التجويع مع تناول الكحول أو بدونسه، علمى بعض المسارات الأيضيه فى الجرذان. تشير النتائج إلى أن التجويسع يسبب إنخفاضاً فى مستوى جلوكوز الدم وكذا تركسيزات البروتينات الكليسة والألبيوميسن، بينما ترداد نسب هرمونسى الإنسمولين والجلوكاجون بالسيرم مقارنة بالمجموعسات الضابطة.

ومن ناحية أخرى، فإن أنشطة إنزيمات السيرم وتشمل : الإنزيمات الناقلة لمجموعات الأمين واللكتات ديهيدروجينيز تزداد إستجابة للتجويع الكامل، وعلمى النقيض من ذلك تتخفض أنشطة إنزيمى الفوسفاتيز القاعدى والجاماجلوتاميل تراسفيريز، بينما لم يتأثر نشاط إنزيم الفوسفاتيز الحامضي تقريباً.

أدى استهلاك الإيثانول إلى إنخف اض أنشطة إنزيم الألانين أمينوتر انسفيريز واللكت ات ديهيدروجينيز، بينما تسبب فى إرتفاع نشاط إنزيم الجاماجلوت اميل تر انسفيريز فسى السميرم فسى الحيوانات المجوعة.

لقد سجل إنخفاض فى محتوى جليكوجين الكبد فى كـل مـن الجـرذان المجوعـة وكـذا التـى تتناول الكحول مع التجويع، كما لوحظ أن أنشسطة الإنزيمـات الناقلـة لمجموعـات الأميسن تنخفسض فى الكبد إستجابة لعملية التجويع، ويزداد نشسـاط إنزيسم السكسسينك ديسهيدروجينيز فـى الكبـد فـى اليوم الأول والثالث من التجويع، بيتما ينخفض فـى اليـوم الخـامس.

لوحظ أن إستهلاك الإيثانول يتسبب فى إرتفاع نشاط إنزيسم الألانيان أمينوتر انسفيريز فى الكبد بينما يؤدى إلى إنخفساض فسى محتوى الجليكوجيس، وفسى أنشطة إنزيمات الاسبارتات أمينوتر انسفيريز والسكسينك ديهيدروجينيز فى أكبساد الجرزان المجوعية.

هذا، وتدعم هذه الدراسة الإفتراضات التي تشمير إلمي أن عمليمة التجويم تحدث إضطراباً فمي مختلف العمليات الأيضية وأن الكحول يؤدي إلى تفساقم هذا الإضطراب.