- Sci. J. Fac. Sci. Menoufia Univ, VoL.VIIII (1993).147-169

EFFECT OF SNA 415, STRAIN OF BACILLUS THURINGIENSIS AND SCHISTOSOME

INFECTION ON THE SURVIVALNESS AND FECNDIUTY OF BIOMPHALARIA ALEXANDRINA SNAILS

G.Y. Osman; A.M.Mohamed, H.I Negm and A.H Mohmed.

Zoology Dept. Fac Science, Menoufia University

Shebin El-Kom, Egypt

ABSTRACT

Effect of SAN415 strain of <u>Bacillus</u> thuringiensis and Schistosome infection on snsil survivalness and fecundity was investigated, in a population of laboratory-bred <u>Biomphalaria alexanderina</u>. Both the survival rates and egg production capacity)assessed by determing number of egg masses / 10 snail / week ; number of eggs / egg mass and histological examination of the ovitestis) have been adversely affected by <u>Schistosoma mansoni</u> miracidial infection. The effect is inversiy proportioal to the number of infecting miracidia and by treatments with a sublethal concentration of SAN 415 strain of <u>Bacollus</u> thuringiensis.

INTRODUCTION

<u>Bacillus</u> thuringiensis. in its commercial bacterial preparation is recently used as a biological agent to control insects (Armstrong

Effect of SAN 415, strain of Bacillus thuringiensis and Schistosome

Mummigatti & Roghunathan, 1988; Nishiura, 1988 and Tottier <u>et</u> <u>al.</u>, 1988); parasitic nematodes (Ignoffo and Dropkin, 1977; Bottjer & Bone, 1987; Osman <u>et al.</u>, 1987 and Osman, 1991) and mloouscs (Terytze & Hogman, 1986; Osman & Mohamed, 1991 and Osman <u>et al.</u>, 1992).Previous trials with <u>B. thuringiensis</u> as a <u>biological</u> agent against the molluscan gastropode snail <u>Biomphalaria alexandrina</u> the vector of Schistosomiasis proved its high potentiality as a molluscicidal agent (Osman and Mohamed, 1991). The comparative molluscicidal action of 4 different preparations of <u>B. thuringiensis</u> was investigated. It has been concoluded that <u>B. thuringiensis</u> in its prparation SAN 415, has proved to be the most potent prepation against <u>B. slexandrina</u> (Osman <u>et al.</u>, 1992) The authers reported that exposure of snails to 500 ppm of SAN 415 for 3 days caused complete sterilization.

Previous investigations have proved that Schistosome infection of snails lead to complete inhibition of the egg laying capacity (Mehleman, 1972; Cheng <u>et al.</u>, 1975; Ishak & Mohamed, 1975 and Mohamed, 1978).

It was proved that develpment of miracidia to mother and daughter sporocysts in the hepatopancreas of the snails was accompaned by releasing toxic material called Schistosomine. This toxine (s) caused inhibition in spermatogenic and oogenic processes (El-Saadany & Mohamed, 1990)

The present work was planned to compare the effect of both

Schistome infection and pathogenic effect of <u>B</u>. <u>thuringiensis</u> on the fecudity of the hermaphrodite snail <u>B</u>. <u>alexandrina</u>. This was followed by histological studies on the hermaphrodite gland of treated snails.

MATERIAL AND METHODS

The snails used in the present study, <u>B</u>. alexandrina were collected from Abou-Rawash, Giza Governorate, Egypt the snails were examined for narural infection with trematode parasites. Negative snails were maintained in a glass aquaria, at room temprature.

All snails were maintained in dechlorinated water uneder standard conditions (photoperiod 16 L / 8 D, water temperature 24-25 C° and PH 7+0.4, Chu et al., 1966 b; 10 snalls per one litre). Snails were fed fresh and dried lettuce leaves added daily.

Laboratory snails were initiated by collecting fresh egg masses laid by wild snails on thin polyethylene sheets or a small pieces of foam placed on the surface of the aquaria (Oliver <u>et al.</u>, 1962). The polyethlene and foam containing egg masses were removed from the aquaria to beakers (250 cc) containing dechlorinated tap water and fragments of chicken egg-schell as a calcium supplement since the local supply of water is soft (Thornhill <u>et al.</u>, 1986). They were examined daily and emerging snsils were trnsfered with a fine brush, few days after hatching, to special rearing aquaria where they were kept until used for studying proposes.

Effect of SAN 415, strain of Bacillus thuringiensis and Schistosome.....

Effect of infection on snail suvivalness and fecundity

Unifected laboratory-bred <u>B</u>. <u>alexandrina</u> snails were classified according to size of juvenile and adult groups 3 & 5 mm respectively) and both were exposed to various numbers of miracidia. The level of miracidial infection were , 4-6 and 8 miracidia for each snail in each experimental group. Survival rates and number of egg masses laid within each snail group were recorded twice a week over a period of eight weeks. A control uninfected group of snails of both sizes were inspected in parallel terms of both paramters.

Effect of SAN 415 Strain of <u>Bacillus</u> thuringiensis on the survivalness and fecundity of snails

The tested commercial bacterial pathogen insecticide <u>Bacillus</u> <u>thuringiensis</u> Berliner was used as SAN 415 strain (32000 1.U. / mg.) A test solution was made up with dchlorinated tap water, PH 7.5-7.7. Stock solution of the experimental material was made on basis of weight volum (O.4 mg / L.) The effect of tested material was investigated for infected and uninfedted adult snails. The tested solution was changed weekly . Control snails maintained under the same experimental conditions were used.

The survival rate and number of egg-masses laid by the snails were recorded daily. Observations were extended over a period of 21 days to determine the effect of prolonged exposure to SAN 415, on the oogenesis and spermatogeesis of <u>B</u>. alexandrina snails. At the end of experimental periods, the ovotestis was dissected out,

fixed in Bouin,s fluid, embedded in paraffin wax and sectioned at 5um. Sections were stained with haematoxylin and eosin and inspected and photographed by light microscopy.

RESULTS

A) Survivalness and fecundity of laboratory - bred snails

The survival rate and fecundity of sexually - mature, adult snails initiated and bred under laboratory conditions, were assessed by determing the number of dead snails, the number of egg masses and the number of eggs in each, twice a week for a period of eight weeks. Data collected from three groups, each consisted of fourty size and age - matched <u>B</u>. <u>alexandrina</u> reared separately and maintained under identical conditions, are given in (Fig. 1), Within the limits of the experimental setting, survival of snails maintained a fairly high rate and yet expressed a slow decline reaching a minimum of 70 % by the end of the experimental period. Over the same observation period, the total reproductive out put of fourty snails was 66 egg masses containing 607 eggs (92 \pm 1.5 eggs / egg mass). The number of egg masses produced per 10 snaols per week (Fig. 1) showed, after an initial unproductive period of two weeks, a gradual increase with time, reaching a peak by fifth week.

Examination of histolgical sections of ovotestes, excised and fixed from about 10 snails at weekly intervals, indicated that the organ is composed of a number of intact acini supported by connec-





Srvival (°) and egg mass production / 10 snails / week (Δ) of normal, mature B. alexandrina. Each point in the cuves is the mean value of three separate series of experiments with SD 5.





mass production / 10 snails / week of 4-6 miracidia- infected (- Δ -) and 8 miracidia- infected (.. Δ ..) mature B. <u>alexandina</u>. Each point in the cuves is the mean value of three separate series of experiments with SD 2.5

tive tissue elements (Fig. 4). In average, each acinus is about 215 um in diameter, lined with germinal epithelium and cxhibits the various developmental stages of male and female sex cells along with the corresponding supporting cells. Male spermtogonia, primary and secondary spermatocytes, spermatids and sperm were generally located in clusters occupying a central position within an acinus (Fig. 5). Scattered along the periphery of the acinus, on the other hand, oogonia, primary and secondary oocytes along with mature ova surrounded by follicular cells, were located. Reflecting the data on egg production in (Fig. 1) the histological appearance of the ovotestis was indicative of active spermatogenesis as well as oogenesis within the group of snails observed.

B) Suvivalness and fecundity of experimentally - infected snails

Groups consisting of fourty age-and size-matched snails were reared speparately, either infected by 4-6 miracida / snail or 8 miracidia/ snail after onset of maturity and inspected for survival rates and fecundity measures over an observation peroid and laboratory conditions identical to groups of uninfected, otherwise matched snails.

As depicted on (Fig 2), and as compared to (Fig. 1), it was evident that the effect of infection of suvivalness and oviposition was dependent on the number of miracidia to which an individual snail was exposed, Although infections with 4-6 mivacidia did vot

alter significantly the suvival rate of snails, infection by 8 miracidia per snail led to the death of about 50 % of snails by four weeks postionfections, yet with no further mortalities recorded thereafter. Over the same observations, period, the total reproductive out put per fourty snails infected with 4-6 miracidia / snail and 8 miracidia / snail was 61 egg masses containing 616 eggs (10.1 ± 1.2 eggs / egg mass) and 19 egg masses containing 116 eggs (6.1 ± 1.3 eggs / egg mass), respectively. It is noteworthy that while among snails exposed to 4-6 miracidia the number of eggs produced per 10 snails per week (Fig. 2) was maintained more or less constant throught the inspection period, the same numbers varied considerably among snalils exposed to 8 miracidia. In addition to the marked redcution in the overall productivity among snails exposed to 8 mracidia, egg prodution showed a gradual increase during the first four weeks post - infection and there after declined shraply and almost disappeared by the end of the observation period.

In a direct correlation to the above observation while ovitestest of snails exposed to 4-6 meracidia were essentially similar to uninfected snails (Fig. 4 &5), the same organ in snails exposed to 8 miracidia was characterized histolgically by the presence of a considerable number of degenerated acini (Fig. 6), Apperantaly, intact acini of reduced diameter (173, 13 ± 20.13 um; 10 snails inspected still however, persisted, and these exhibited signs of marked reduction in sperm production as well as complete inhibition of oogenesis, which was represented by few scattered degenerative oocytes.





(Figure 3) Survival of SAN 415 treated, uninfected (Ø) and 4-6 miracidia-infected (..Ø..) mature B. alexandrina. Also shown is the egg mass production / snail / week of SAN 415 treated, uninfected (- Δ -) and 4-6 miracidia - infected (.. Δ ..) snails. Each point in the curves is the mean value of three seperate series of experiments with SD 4.5.



effect of SAN 415, strain of Bacillus thuringiensis and Schistosome ...

EXPLANTION OF FIGRUES

Figure 4 & 5

Light micrograph of an eosin - haematoxlin- stained normal ovotestis of mature snails showing acini (Ac) of normal size (4, x250) and active spermatogenesis and oogenesis (5, x400). S, sperm; Sc, spermartocyte; O, ovum; Oc, oocyte; F, follecular cells, Bar = 200 um.

Figure 6

Light micrograph of an eosin - haematoxlin - stained ovotestis of 8 miracidia- infected, mature snails showing acini (AC) of rduced size, lack of oogensis and markedly reduced spermatogenesis. Spg, stages of spermatogenesis. x400.

Figre 7 & 8

Light micrograph of eosin - haematoxlin - stained ovotestes of SAN 415 - treated, uninfected (7) and 4-6 miracidia - infected (8), mature snails showing acini (Ac) of reduced size, lack of oogensis and abrogation of spermatogenesis. Isp, irregrlar sperm; Spg, stages of spermatogenesis. x400. Effect of SAN 415, strain of Bacillus thuringiensis and Schistosome

C) Survivalness and fecundity of infected and SAN 415 treated snails

Snails treated with SAN 415 and then exposed to 4-6 miracidia / snail, since by the first week about 75 % of snails survived these treatments and thereafter started to decline, so that by the third week no survivors were observed (Fig. 3), On the other hand, a slower decline in survival rates was observed among snails treated with SAN 415 and left uninfected, since after an initial 25 % mortalities in the first week, additional 40 % mortalities were recorded by the end of four weeks.

At any given time of the observation period, snails treated with SAN 415 exhibited no oviposition activities, and thus was true in either uninfected or in snails infected with 4-6 miracidia / snail. Inspected histologically, ovotestes of uninfected snails or snails exposed to 4-6 miracidia / snail equally expressed signs of degneration. In both snail grorps, the acini were of reduce diameter $(142.8\pm 18.2 \text{ um}, 10 \text{ snails inspected or totally disrupted, spermato$ genesis represented by clusters of spermatozoa whereas otherstages were less prominent and ooginesis totally inhibited and represented by denucleated oocytes of unedfined shape and reducedsize (Fig. 7 & 8).

DISCUSSTION

Considering several aspects of snail physiology as growth, fecundity and mortality, data based on field population have long suf-

fered uncertainties due to seasonal fluctuations, random infections as well as variations in climatic factors (Anderson et al;, 1982; woolhouse, 1989). In the present study, these variations have been rendered largly uneffective in populations of laboratory bred snails, which were initiated and maingained according to standard procedures (Thornhill et al ., 1986) Results recorded here regarding survival rates, egg-laying capacity as well as the integrity of the reprocuctive organs in uninfected mature B. alexandrina are in accordance with abservation reported for <u>B</u>. glabrata (Minchella, 1985), B. pfeifferi (Mrkanga, 1981), Lymnaea catascopium (Loker, 1979) and Lymnaea stagnalis (Sluiters et al ., 1980) reared under comparable conditions. Interestingly, S. mansoni infections seemed to be well tolerated by this population of laboratory - conditioned \underline{B} . alexandrina. since over 70 % of snails originally exposed to 4-6 miracidia each, were still alive eight weeks post exposure. Increasing the exposure dosage from 4-6 to 8 miracidia per snail, subsequently decreased survivorship. yet still 50 % of the snails were survived throughout the same observation period. In relation to this observation, several investigators have reported that Schistosome infections adversely affect the survival of the molluscan host (Chu et al ., 1966 a; Sturrock, 1970; Lo, 1972; Meuliman, 1972; Mohamed and Ishak 1982), although others observed increased survival in infected snails (MsClelland and Bourns, 1969). These conflicting obser-

vations are directly dependent upon variations in the host-parasite combination studied and may prove to be linked to specific host genetic counter adaptation mechanisms to parasitism (Minchella,

Effect of SAN 415, strain of Bacllus thuringiensis and Schistosome

1985).

Another physiological measure that is repeatedly reported to be modulated by Schistosome infection, is fecundity. In respones to infection, sharp reduction and eventual cessation of egg production among infected snails in one hand, has been a widely accepted observation (Meuleman, 1972; Baudoin, 1975; Mohamed and El Fiki, 1980). On the other hand, there has been evidence of a significant increase in egg-laying in either mature or immature infected snails (Minchella, 1985; Thornhill et al., 1986). Two brust of egg-laying immmedately following parasite exposure has been interpreted as a mlluscan adaptation in respones to trematode parasitism, and has been termed fecundity compensation (Minchella and Loverde, 1981). AN observation that could be correlated to this adaptation phenomenon was obtained in the present study. Mature snails infected with 4-6 miracidia seemed, in terms of fecudity measures to tolerate such infection and produced an overall number of egg masses and eggs per egg mass comparable to uninfected controls, yet without a pronounced burst in egg-laying at any given time post infection. Increasing the miracidial load, both egg production activity as well as the integrity of the ovotestis were markedly affected, suggesting that fecundity in <u>B</u>. alexandrina may after all be inversely proportional to the number of infecting miracidia as observed earlier with other related moullusks (Pan, 1965; Loker, 1979; Makanga. 1981; Crews and Each, 1986)

In addition to <u>S</u>. mansoni miracidia which represent (natural)

burdens on snail survivalness and fecundity, the effect of a sublethal concentration of a SAN 415 strain of the bacterium B. thuringiensis on theses parameters was investigated. Although previous reports screening natural bacterial pathogens among moribund snails have ruled out the presence of such bacterium (Cheng, 1986), its adverse effect on survivalness was remininscent of heavy natural infection. Interestingly treatment of either uninfected or infected snails have also resulted in complete abrogation of egg proeduction and yielded what seamed to be a state of castration. Total cessation of egg laying as early as two days posttreatment was paralleled by elevated sings of atrophy in the ovotestis, where retaradtion of spermatogensis and a complete absence of oogenesis was apparent. To our knowledge snail castration has been always attributed to infection by direct mechanical bolckage of neutrient transport due to the pressure exerted on gonadal tubules by invasive larval stages (James, 1965), or indirectly via the active secretion of cytolytic chemicals of hormones by the developing larvae (Chen 1983), In this study, the observation that no sporocyts were detected in the gonadal tissue of infected snails, farvous the suggestion that in <u>B</u>. alexandrina, gonadal elements may be selectively -susceptible targets to chemical modulations. In uninfected snails, these modulations seemed to be mimced in resposne to sublethal dose of Bacillus treatments indicating that possibly other toxic compounds may have similar effects. Although these suggestions demand futher investigations, our findings may have important implications in the biological control of <u>B. alexandrina</u>.

Effect of SAN 415, strain of BacIlus thuringiensis and Schistosome

REFERENCES

- Anderson, R.M ; Mercer , J. G.; Wilson , R.R. and Carter, N.P. (1982) Trnsmission of <u>Schistosoma mansoni</u> from man to snail : Experimental studies of miracidial survival and infectivity in relation to larval age, water temperature, host size and host age. parasitology, 85: 339-360.
- Armstrong, J.I.; Rohrman, G.F. and Beaudreau, G.S. (1985). Delta endotoxin of <u>Bacillus thuringiensis</u> subsp. israelensis. J. Bacteriol., 161: 39-49.
- Baudoin, M. (1975) . Host castration as a parasitic strategy. Evolution. 29: 335-352.
- Bottjer, K.P. and Bone, L.W. (1987), (Changes in morphology of <u>Trichostrongly colubriformis</u> eggs and Juveniles caused by <u>Bacillus thuringiensis</u> israelensis. J. Nematolgy, 19 (3): 282-286.
- Cheng, T.C. (1983). Internal defense mechanisms of molluscs against invading microorganisms: Personal reminiscences. Trans. Amer Microsc. Soc., 102: 185-193.
- Cheng. T.C. (1986). Biological control studies : Bacteria Associated with Moribund <u>Biomphalaria glabrata</u> (Mollusca) in the Laboratory. Journal of Invertdbrate Pathology ., 47: 219-224.
- Cheng. T.C.; Rodrick, G.E.; Foley, D.A and Koehler, S.A. (1975). Release of Lysozyme from hemolymph cells of <u>Mercenaria mercenaria</u> during phagocytosis . J. Invertebr. Path-

ol., 25: 261-265

- Chu, K.Y.: Sabbaghian, H. and Massoud, J. (1966 a). Hostparassite relationship of <u>Bulinus trancatus</u> and <u>Schistosoma haematobium</u> in Iran. 2. Effect of exposure dosage of miracidia on the bilology of the snail host and the development of the parasites. Bull. WHO., 34: 121-130.
- Chu, K.Y.; Massoud, J. and Sabbaghian, H. (1966b) Host parasite relationship of <u>Bulinus trancatus</u> and <u>Schistosoma hae-</u> <u>matobium</u> in Iran. 3. Effect of water temperature on the ability of miracidia to infect snails. Bull. WHO., 34: 131-133.
- Crews. A.E. and Esch, G.w. (1986). Seasonal dynamics of <u>Halipe-gus</u> occidualis (Trematoda, Hemiuridae in Helisoma anceps and its impact on fecundity of the snail host. J. Parasitol., 72:115-121.
- Davidson, E.W. and yamamoto, T. (1985) Isolation and assay of the toxin components from the crystals of <u>Bacillus thu-</u> <u>ringiensis</u> israelensis Curr. Microbiol., 11; 171-174.
- El-Saadny, M.M. and Mohamed, A.M (1990). Effect of Trichobilharzia <u>ocellata</u> and <u>Schistosoma mansoni</u> infection on the ultrastructure of the albume gland of their snail host <u>Lymnae stagnalis</u> and <u>Biomphalaria alexandrina</u> respectively. Folia Morphologica
- Ignoffo, C.M. and Dropkin, V.H. (1977). Delaterious effects of the therostable toxin of <u>Bacillus thruingiensis</u> on species of

Effect of SAN 415, strain of Bacllus thuringiensis and Schistosome....

soil inhabiting, myceliophagous, and plant-parasitic nematodes. J. Kans. Entomol. Soc. 50: 394-398.

Ishak, M.M and Mohamed, A.M. (1975). Carbohydrate metabolism in uninfected and trematode infected <u>Biomphalaria</u> <u>alexandrina</u> and <u>Bulinus truncatus</u>. Comp. Biochem. physiol. 218: 499-505.

- James, B.L (1965). The effects of parasitism by larval digenea on the digestive gland of the interidal prosobranch <u>Literorina Saxatilis</u> subsp. <u>tenebrosa</u> (Mont.).J. Nat. Hist. 2:21-37.
- Lo, C.T. (1972). Compatibility and host-parasite relationship between species of genus <u>Bulinus</u> (Basommatophora: Planorbidae) and an Egyptian strain of <u>Schistosoma hae-</u> <u>matobium</u> (Trematoda : Dignea) Malacologia., 11:225-280.
- Loker. E.S. (1979). Pathology and host responses induced by <u>Schistosomatium douthitti</u> in the fresh water snail <u>Lym-</u> <u>naea catascopium</u>, J. Invertebr. Pathol., 33;265-273.
- Makanga, B. (1981). The effect of varying the number of <u>Schistos-</u> <u>ma mansoni</u> miracidia on the reperoduction and suvival of <u>Biomphalaria pfeifferi</u>. J.Invertebr. Pathol., 37: 7-10.
- Mcclelland, G. and Bouns, T.K.R (1969). Effects of <u>Trichobilharzia</u> <u>ocellata</u> on growth, reprodution, and suvival of <u>Lymnaea</u> <u>stagnalis</u>. Exper. Parsit., 24:137-146.

- Meuleman, E.A. (1972) .Host -parasite interrelationships between the freshwater pulmonate <u>Biomphalaria pfeifferi</u> and the trematode <u>Schistosoma mansoni</u>. Neth. J. Zool., 22;355-427.
- Minchilla, D.J. (1985). Host life-history variation in response to parasitism. parasit., 90:205-216.
- Minchella, D.J. and Loverde, P.T. (1981). A cost of increased early reprocuctive effort in the snail <u>Biomphalaria glabrata</u>. The American Naturalist., 118: 876-881.
- Mohamed, A.M. (1978). Effect of <u>Trichobilhrzia ocllata</u> infection on growth rate and end products of the anaerobic metabolism in hermaphrodite pulmonata snails <u>Lymnaea stagnalis</u>, Egyptian J. Bilh. 4,No.2, 187-194.
- Mohamed, A.M. and El-Fiki, S.A. (1980). Egg procuction and tissue glycogen of different age groups of <u>Biomphalaria</u> <u>alexandrina</u> infected with <u>Schistosoma mansoni</u>, J. Egypt Soc. parasit., 10:65-72.
- Mohamed, A.M. and Ishak, M.M.(1982). Comparative effects of Schistsome infection and starvation on the respiratory transport chain of the snails <u>Biomphararia alexandrina</u> and <u>Bulinus truncatus</u>. Comp. Biochem. and physiology. 71B., 289-298.

Effect of SAN 415, strain of Bacllus thuringiensis and Schistosome

Mummigatti, S.G.and Raghunathan, A.N (1988) Production of <u>Ba-</u> <u>cillus thuringiensis</u> Var Kurstaki by three different methods and its relative toxicity to <u>Bombyx mori</u>. J. of Invertebrate Pathology 51: 115-118.

- Nishiura, J.T. (1988). Fraction of two mosquitocidal activities from Alkali-solubilized extracts of <u>Bacillus thuringiensis</u> subspecies israelensis spores and parasporal inculusions. J. Invertebr. Pathol., 51: 15-22.
- Noble, E.R. and Noble, G.A. (1971). Parasitology: The Biology of Animal Parsites lea and Febiger. Philadlphia.
- Oliver, L.; Hsakins, W.T., and Gruian, J. (1962). Action of very low concentration of Na Pentachlorophenate on freshly laid eggs of <u>Australorbis glabratus</u>. Buol. Wld. Hith. Org., 27: 87-94
- Osborne, L.S.; Bouclas, D.G. and Lindguist, R.K. (1985). Activity of Bacillus thuringiensis Var. israelensis on Eradysia coprophila (Diptera: Seiaridae). J. Econ Entom. 78 (4): 922-925.
- Osman, G.Y. (1991). Effect of soil solarization and Bacillus <u>thurn-</u> <u>gensis</u> berliner on <u>Rotylenchulus reniformis</u> and <u>Meloi-</u> <u>dogyne incognita</u> (Tylenchida; Nematoda). J. Egypt. Ger Soc. Zool. Vol (3) 85-91.
- Osman, G.Y. and Mohamed, A.M. (1991). Bioefficacy of bacterial insecticide, <u>Bacillus thuringiensis</u> Berl. as biological

control agent against snails vectors of Schistosomiasis in Egypt . Anz Schadlingskde., Prlanzenschutz, Umweltschutz, 64, 136-139

Osman, G.Y., Mohamed, A.M. and Jamel Al-Layl,, K.(1992). Ubberdie molluscizede Wifkung von <u>Bacillus thuringiensis</u>praparaten zur bologischen Bekampfung von <u>Biomphalaria</u> <u>alexandrina</u> Schnecken, den Zwischenwirten des Bilharziose-Saugwurms <u>Schistosoma mansoni</u>. Anz. Scadoingskde., Pflanzenschutz, Umweltscutz 65: 67-70.

- Osman, G.Y.; Salem, F.M and Ghattas, A. (1987). Biofficacy of two bacterial insecticide strains of <u>Bacillus thuringiensis</u> as a biological control agent in comparison with a Nematicide, phenamiphos, on certain parasitic nimatodes. Anz. Schadlingskde, pflanzenschutz, Umwelschutz 61: 35-37.
- pan, C. (1965). Studies on the host-parasite relationships between <u>Schistosoma masoni</u> and the snail <u>Australorbis glabratus</u>. Amer J. Trop. Med Hyp., 14:931-976.
- Slutiers, J.F.; Brussaard-Wust, C.M. and Meuleman, E.A. (1980). The relationship between miracidial dose, production of cercariae, and reproductive activity of hte host in the combination <u>Trichobilharzia ocellata</u> and <u>Lymnaea stagnalis</u>. Zeitschrift fur Parasitenkunde., 63: 13-26.

Sturrock, B.M. and Sturrock, R.F. (1970). Laboratory studies of the hostparasite relationship of <u>Schistosoma mansoni</u> and <u>Bi-</u>

Effect of SAN 415, strain of Bacllus thuringiensis and Schistosome

omphalaria glabratus from Stlucia, West Indies. Ann. Trop. Med Parasitol., 64: 357-363.

- Terytze, K. and Hofmann, G. (1986). Die Wikung von Bakterienprapraten (<u>Becillus thuringiensis</u> Berliner) Zur von Nscktschnecken in Gerbera- Bestanden. Arch. phytopathol. pflanzenschutz, Berlin., 22: 361-363.
- Thornhill, J.A.; Jones, J.T. and Kusel, J.R. (1986). Increased oviposition and growth in immature <u>Biomphalaria globrata</u> after exposure to <u>Schistosoma mansoni</u>. Parasit., 93: 443-450.
- Tottier, M.R. Mooris, O.N. and Dulmage, H.T. (1988). Susceptibility of the Bertha Armyworm, <u>Mamestra Configurata</u> (configurata (Epidoptera, Noctuidae) to sixty -one strains from ten varieties of <u>Bacillus thuringiensis</u> j. of Invertebrate pathology, 51:242-249.
- Wool House, M.E.J. (1989). On the interpretation of ageprevalence curves for Schistosome infection of host snails. Parasit.99: 47-56.

تأثير البكتريا باسلس ثيرينجينسس سلالة سان١٥ ع والعدوى بالبلهارسياعلى بقاء وخصوبة قواقع بيومغلاريا الكسندرينا

جمالات یوسف عثمان محمد احمد مصطغی هدی ابراهیم زجم عزة دسن محمد

قسم علم الديوان – كلية العلوم – جامعة المنوفية – شبين الكوم

تم دراسة تأثير كل من البكتيريا باسلس ثيرنجينسس سلاله سان ٤١٥ والعدوى بميراسيديا الشستوسوما مانسونى علي معدل بقاء وخصوبة السلالة المعملية لقوقع بيومفلاريا الكسندرينا أوضحت النتائج مايلى:

أن كل من معدل البقاء على الحياة وكفاءة إنتاج البيض (والتى حددت بمعدل وضع القوقع لكتل البيض وعدد البيض فى كتلة البيض الواحدة) وكذا دراسة الفحص النسيجى للمنسل الخنثوى للقوقع تتأثر بعدوى القوقع بميراسيديا الشستوسوما والتأثير هذا يتناسب تناسباً عكسياً مع كل من عدد الميراسيديا والمعاملة بالتركيز تحت الميت من بكتريا باسلس ثيرنجينسس سلالة سان ٤١٥.